

Method Development and Validation of Rifampicin and Isoniazid & Pyrazinamide and Ethambutol Hydrochloride by Using RPHPCL Method

R.Nillimarani

M.Pharmacy,

**Sri Vani School of Pharmacy Chevuturu,
Krishna (Dt), G.Koundru (Md)-521229, India.**

Mr. G.Visweswara Rao

Associate Professor,

**Sri Vani School of Pharmacy Chevuturu,
Krishna (Dt), G.Koundru (Md)-521229, India.**

Y.Chaitanya Kumar

Assistant Professor,

**Sri Vani School of Pharmacy Chevuturu,
Krishna (Dt), G.Koundru (Md)-521229, India.**

Dr. K.Narendra Kumar Reddy

Professor,

**Sri Vani School of Pharmacy Chevuturu,
Krishna (Dt), G.Koundru (Md)-521229, India.**

A new simple, rapid and sensitive reversed-phase high-performance liquid chromatography method was developed and validated as per International Conference on Harmonization guidelines, Q2 (R1), for simultaneous estimation of isoniazid, pyrazinamide and rifampicin in solid lipid nanoparticles. Separation was achieved on a 250×4.6 mm, 5 µm, C-18 column using a linear gradient flow rate of 1.5 ml/min. Isoniazid, pyrazinamide and rifampicin were identified based on their retention times as compared to standards and confirmed with characteristic spectra on a spectrophotometer at 238 nm and eluted at 3.787, 4.173 and 11.273 min, respectively, achieved within 20 min. This method was linear, precise with % RSD values of 0.18 % for isoniazid, 0.15 % for pyrazinamide and 0.47 % for rifampicin, accurate with mean recovery yields of 101.312 % for isoniazid, 99.910 % for pyrazinamide and 99.767 % for rifampicin and selective over the concentration range of 10-150 % for all the three drugs. This method is suitable due to its simplicity and accuracy for routine quality control and stability analysis of antitubercular drugs-loaded solid lipid nanoparticles. Key words: Simultaneous estimation, RP-HPLC, isoniazid, pyrazinamide, rifampicin, solid lipid nanoparticles. Tuberculosis (TB) treatment regimens followed in India under the Revised National Tuberculosis Control Programme (RNTCP) 1997.

The initial phase of category-III TB (new cases of smear-negative pulmonary TB with limited parenchymal involvement or less severe forms of extrapulmonary TB, viz., lymph node TB, unilateral pleural effusion, bone (excluding spine), peripheral joint or skin TB) treatment, the combinations of first-line antitubercular drugs (ATDs), isoniazid (INH), pyrazinamide (PYZ) and rifampicin (RIF) are widely used daily per week for at least 2 mo[1]. Structures of these first-line three ATDs are depicted in fig. 1A, B and C, respectively. These combination preparations are available in the world market. The International Union against tuberculosis and lung disease and the World Health Organization recognize the potential benefits of using such preparations. Spectrophotometric and reversed-phase highperformance liquid chromatography (RP-HPLC) techniques have been reported for individually or the combination of two or more ATDs and other drugs estimation in biological samples and pharmaceutical formulations[4-19]. In the previous years, simultaneous estimation of INH, PYZ and RIF in fixed combination of unit doses by

Cite this article as: R.Nillimarani, Y.Chaitanya Kumar, Mr. G.Visweswara Rao & Dr.K.Narendra Kumar Reddy, "Method Development and Validation of Rifampicin and Isoniazid & Pyrazinamide and Ethambutol Hydrochloride by Using RPHPCL Method", International Journal & Magazine of Engineering, Technology, Management and Research, Volume 5, Issue 12, 2018, Page 114-119.

highperformance thin layer chromatography[20], multivariate spectrophotometric calibration method [21] and calorimetric analysis[22] have been published. However, many of these methods suffer from limitations such as complex and tedious procedures and are not validated as well. HPLC is the method of choice for analysis of multicomponent pharmaceutical preparation because of its sensitivity, reproducibility and specificity. The problems associated with optimization of chromatographic conditions such as selection of column type, column temperature, the composition of the mobile phase, selection of the specific wavelength and injection volume. In spite of the fact, this method undoubtedly provides the more sensitive determination than the spectrophotometric methods.

It has now become the method of choice for most of the drug and their combinations. Gaitonde and Pathak reported a RP-HPLC method for the simultaneous estimation of INH, PYZ and RIF. They used tetrabutylammonium hydroxide as an ion-pairing agent in the mobile phase which shortens column life[23]. Calleri et al. developed an HPLC method having high buffer concentration in the mobile phase of acetonitrile (A) and 50 mM phosphate buffer pH 3.5 (B). The gradient profile was (A:B) 3:97 v/v for 5 min, then a linear gradient to 50:50 v/v at 30 min and return to 3:97 v/v in 10 min. Chromatography was performed at room temperature using a flow rate of 1 ml/min and a run time of 40 min.

Diode-array detector with three different corresponding maximum of 254 nm for RIF, 261 nm for INH and 265 nm for PYZ were used, this method was time consuming and conditions specific[24]. Dhal and Sharma reported RP-HPLC method for simultaneous estimation of pyridoxine hydrochloride, INH, PYZ and RIF. The result showed that the highest values for limits of detection (LOD) were 0.043, 0.063, 0.036 and 0.059 µg/ml and limits of quantification (LOQ) were 0.13, 0.19, 0.11 and 0.18 µg/ml, respectively.

So, this method is not sensitive to detect the lowest concentration of combined ATDs[25]. The Indian Pharmacopoeia 2014 and United States Pharmacopoeia 2016 published HPLC method for simultaneous determination of INH, PYZ and RIF in the tablet and capsule dosage forms[26,27]. The aim of this study was to develop and validate an accurate and sensitive RP-HPLC method for simultaneous estimation of INH, PYZ and RIF in solid lipid nanoparticles (SLNs). The validation results obtained showed that this method was simple, accurate, precise and suitable for the determination of ATDs in SLNs.

MATERIALS AND METHODS

The working standards employed for INH, PYZ and RIF were obtained from Indian Pharmacopoeia Commission (IPC) Ghaziabad, Uttar Pradesh, India. The Poloxamer 118 (Pluronic F-68) was purchased from HiMedia Laboratory Pvt. Ltd., India. Sodium taurocholate was purchased from LOBA Chemie, India. Stearic acid (octadecanoic acid), mannitol was purchased from Qualigens Fine Chemicals (Division of GSK Pharmaceutical Ltd.). Ortho-phosphoric acid, sodium hydroxide, methanol and acetonitrile of analytical grade were purchased from Thermo Fisher Scientific India Pvt. Ltd. Water was deionized by passing through a Milli-Q™ Integral-3 water purification system (M-Millipore, Fisher Scientific India Pvt. Ltd.). Chromatographic experiments were performed using a HPLC instrument (Agilent Technologies 1260 Infinity with EZ Chrome Elite software) equipped with autosampler, ultraviolet/photodiode-array detector (PDA) and stainless steel C-18 column (Nucleodur®) packed with octadecylsilane bonded to porous silica (5 µm), 250×4.6 mm (Macherey-Nagel GmbH and Co, Düren, Germany) and the column temperature was maintained at 30°.

Chromatographic conditions:

Chromatographic analysis were performed in a linear gradient programme using mobile phase A and mobile phase B with the flow rate of 1.5 ml/min (Table 1).

Chromatographic system set the spectrophotometer at 238 nm and the column temperature was maintained at 30°. The samples were diluted with orthophosphoric acid (OPA) buffer solution pH 6.8±0.02 and 20 µl was injected into the column.

Preparation of the solutions:

Buffer solution was prepared by diluting 1 ml of OPA in 1000 ml of water and adjusts the pH 6.8±0.02 with dilute solution of sodium hydroxide. This buffer solution was equivalent to 17.85 mmol OPA buffer solution. Mobile phase A used was a mixture of 96 volumes of buffer and 4 volumes of acetonitrile. Mobile phase B used was a mixture of 45 volumes of buffer and 55 volumes of acetonitrile.

Preparation of stock and standard solution:

Stock solution was prepared using 0.04 % w/v of INH, 0.2 % w/v of PYZ and 0.08 % w/v of RIF in methanol. The prepared stock solution was stored at 4° until before use. 5 ml of stock solution was transferred to a volumetric flask and diluted up to 25 ml with buffer. The prepared final concentration of INH, PYZ and RIF were 80, 400 and 160 µg/ml, respectively.

Calibration solutions:

Appropriate dilutions of the stock solution were prepared with buffer in 25-ml volumetric flasks. The diluted quality control samples (QCS) of 10, 20, 30,

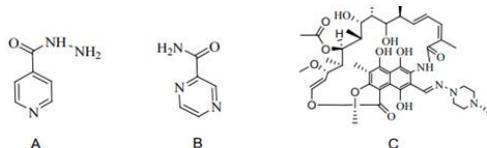


Fig. 1: Chemical structures (A) INH; (B) PYZ and (C) RIF

TABLE 1: THE CONDITIONS OF LINEAR GRADIENT PROGRAMME

Time (min)	Flow rate (ml/min)	Mobile phase A (% v/v)	Mobile phase B (% v/v)
0	1.5	100	0
10	1.5	100	0
11	1.5	0	100
15	1.5	0	100
16	1.5	100	0
20	1.5	100	0

40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140 and 150 % were prepared and analysed for linearity.

Method validation: Validation of the RP-HPLC method was performed as per the International Conference on Harmonization guideline, Q2 (R1)

specifications after the establishment of chromatographic and the experimental conditions [28].

System suitability: To ensure the validity of the analytical procedure, a system suitability test (SST) was established. Percent relative standard deviation (% RSD) of the area, RSD of retention time (RT), USP tailing factor, theoretical plates, and resolution were selected for SST. These parameters were analysed by injecting the standard solution six times. The system suitability results are summarized in Table 2 and could be observed as the parameters analysed were in accordance with acceptance criteria.

Specificity: Specificity of the method was established to ensure that the components of the SLNs dispersion such as lipid(s) and surfactant(s), does not interfere in the quantification of the drugs[29]. The specificity of the method was evaluated by comparing the chromatograms of ATDs extracted from SLNs and of blank nanoparticles to determine the peak purity.

Linearity: For linearity, stock solutions of INH, PYZ and RIF at 15 concentrations between 10 to 150 % were injected in triplicate. Calibration curves of standard INH, PYZ and RIF were generated by plotting the analyte peak area versus % drug concentration. Linearity is confirmed if the % RSD values of the slope and the intercept are Linearity is confirmed if the % RSD values of the slope and the intercept are <1%.

Precision: The system precision of the method was determined by performing intra-day and inter-day analysis by triplicate injections of the QCS. The % RSD (% RSD=SD/mean×100) and % accuracy (% accuracy = mean area of test/mean area of standard×100) were calculated by comparing the theoretical and measured concentration for each case (Table 3). According to the ICH guideline, coefficients of variation for the precision shall not exceed 5 %.

Accuracy: To determine the accuracy of the proposed method, recovery studies were conducted. The extraction process of the method was quantified by comparing three separately extracted sample preparation (SLNs containing ATDs and excipients) against a standard solution. The acceptance criterion for the extraction study was the mean recovery of the extracted ATDs from SLNs relative to the standard solution shown in Table 4.

LOD and lower LOQ: LOD was the lowest concentration of the analyte in a sample that could be detected under the stated experimental condition. LOQ is the lowest concentration of the active ingredients in a sample that could be determined with acceptable precision and accuracy. According to the ICH recommendation, the approach based on the standard deviation (SD) of the response and slope (m) was used for determining the detection and quantitation limits. LOD can be calculated according to the formula, $LOD=3.3 (SD/m)$ and LOQ according to the formula, $LOQ=10 (SD/m)$.

TABLE 2: SYSTEM SUITABILITY RESULTS

Parameters	Limits	ATDs		
		INH	PYZ	RIF
% RSD of the area	Less than 2	0.05	0.05	0.06
RSD of retention time	Less than 1.0	0.09	0.08	0.02
USP tailing factor	Less than 2	1.26±0.069	1.25±0.147	1.27±0.070
Theoretical plates	More than 2000	27647±226.57	33317±267.23	98821±268.59
Resolution	More than 2	-	3.94±0.01	58.72±0.12

System suitability was checked by injecting the working solution six times

TABLE 3: INTRA-DAY AND INTER-DAY PRECISION AND ACCURACY OF INH, PYZ AND RIF

QCS	Amount (µg/ml)			Standard (Intra-days)									
	INH	PYZ	RIF	Found (µg/ml)			Accuracy (%)			Precision (% RSD)			
				INH	PYZ	RIF	INH	PYZ	RIF	INH	PYZ	RIF	
10 %	8	40	16	7.90	39.60	15.90	98.72	99.00	99.35	0.04	0.05	0.47	
50 %	40	200	80	39.64	198.19	79.29	99.09	99.09	99.11	0.18	0.15	0.16	
100 %	80	400	160	79.36	396.56	158.27	99.20	99.14	98.92	0.03	0.07	0.06	
150 %	120	600	240	119.17	595.15	237.53	99.31	99.19	98.97	0.05	0.02	0.08	
				Standard (inter-days)									
10 %	8	40	16	7.85	40.11	15.73	98.09	100.28	98.30	0.14	0.01	0.32	
50 %	40	200	80	39.84	200.36	79.73	99.60	100.18	99.67	0.03	0.05	0.06	
100 %	80	400	160	79.98	400.91	159.74	99.98	100.23	99.84	0.08	0.05	0.07	
150 %	120	600	240	119.98	601.05	240.64	99.98	100.18	100.27	0.01	0.01	0.01	

TABLE 4: RECOVERY ANALYSIS OF ATDs-LOADED SLNS

ATDs	Assay *			Recovery *		
	Found (µg/ml)	% RSD	Mean recovery (%)	±SD	% RSD	% RSD
INH	0.3424	0.53	101.312	939.74	6.11	
PYZ	1.4623	0.06	99.910	188.80	0.12	
RIF	1.1194	0.39	99.767	7.07	0.07	

*Mean of two extractions (each analyzed in duplicate)

Determination of ATDs encapsulation efficiency (EE) in SLNs:

SLNs were prepared using a modified microemulsion technique, freeze-dried and stored at 4°[31]. The mean particle size and the polydispersity index (PDI) were determined using dynamic light scattering (Malvern Instruments Ltd. Zetasizer Ver. 7.11), at room temperature.

To determine the amount of ATDs in SLNs, 5 mg of lyophilized SLNs were transferred into a 5-ml volumetric flask and 3 ml of ethanol was added. This mixture was heated to 50° with sonication and final volume was made up to 5 ml of ethanol[32,33]. The above solution (1 ml) was placed in a 50 ml volumetric flask and diluted with buffer. The sample was ultrasonicated for 10 min and filtered with nylon syringe filter (0.22 µm). Further 1 ml of the filtrate was diluted up to 10 ml with buffer. The sample was analysed by using the designed chromatographic conditions as discussed earlier. Subsequently, the EE was calculated with the help of the reported Eqn.[34], $\% EE = (total\ drug - free\ drug) / (total\ drug) \times 100$. Where, total drug corresponds to the amount of ATDs initially added to the SLNs dispersion and free drug is the amount of the drug not incorporated into the lipid nanoparticle, quantified by RP-HPLC method. The % EE was quantified by RP-HPLC method and the value of % EE was expressed as mean and SD.

RESULT AND DISCUSSION

In order to optimize good separation between all the three ATDs drugs at different buffer pH and organic solvents like acetonitrile and water tested binary and tertiary eluents. Therefore, best chromatographic conditions concluded with different ratio of 17.85 mmol OPA buffer (pH 6.8) and acetonitrile, flow rate of 1.5 ml/min and detected at 238 nm. The analytical profile obtained by injection of a standard working solution was reported in fig. 2. The analytical profiles obtained by injection of extracted spiked SLNs without active pharmaceutical ingredients and of extracted spiked ATDs-loaded SLNs were reported in fig. 3. The SST parameters such as RT, USP tailing, theoretical plates and resolution for optimizing standard chromatogram are tabulated in Table 2. The SST is an integrated part of the analytical method and it ascertains the suitability and effectiveness of the operating system. The results were within the limit and are presented in Table 2 and system suitability chromatogram in fig. 2.

The USP tailing factor for the analytes peaks were less than 2.0, the % RSD of the peak areas responses less than 2 % and the migration times within seconds. Specificity is the ability of the method to measure the analyte response in the presence of its potential impurities and degradation products[35]. The representative chromatograms of INH, PYZ and RIF were identified with the RT as compared with the standard at 3.887, 4.200 and 11.327 min, respectively. The specificity studies revealed the absence of any other excipients interference, since none of the peaks appeared at the same RT, as shown in fig. 3. The specificity was justified from complete separation of drugs from SLNs and indicated that there was no interference in the quantitative determination of ATDs from SLNs components. Linear responses were observed in the range of 8-120, 40-600 and 16-240 µg/ml for INH, PYZ and RIF, respectively. The linear regression analysis obtained by plotting the peak areas of the three analytes versus percent concentration. The result showed excellent correlation coefficients ($R^2 \geq 0.999$) and the linearity data was reported in Table 5.

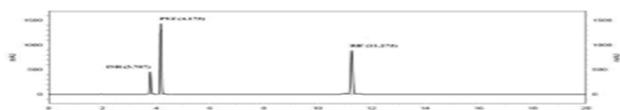


Fig. 2: System suitability chromatogram of INH, PYZ and RIF

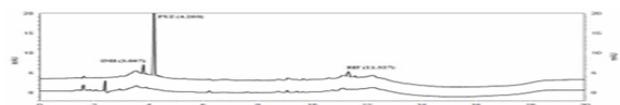


Fig. 3: Analytical profile of extracted spiked SLNs without ATDs and with ATDs
The analytical profile obtained by injection of extracted spiked SLNs without APIs and with ATDs-loaded SLNs

The mean recovery data obtained were within 2 %. The mean recovery yields were 101.312, 99.910 and 99.767 % for INH, PYZ and RIF, respectively. The recovery data is reported in Table 4. Since the results obtained were within the acceptable ± 3 % range, the method was deemed to be accurate. Estimation of LOD and LOQ considered the acceptable SD of the response and the slope of the standard curve. The LOD and LOQ was calculated as 0.0042 and 0.0127 for INH, 0.0023 and 0.0070 for PYZ and 0.0475 and 0.1439 µg/ml for RIF, respectively (Table 6). The SLNs containing ATDs were successfully obtained by microemulsion technique and proposed analytical

method was applied to evaluate the EE of ATDs in SLNs. As shown in the specificity test, no alterations or unusual peaks were observed in the chromatograms during the drug quantitation. The mean recovery yields were 101.312, 99.910 and 99.767 % for INH, PYZ and RIF, respectively. The results demonstrated that the method used was efficient in associating the drugs with SLNs. The average diameter of the solid nanoparticles was 374.1 ± 3 nm and PDI (0.436 ± 0.005) indicating mono-modal size distribution. Therefore, the proposed RP-HPLC method represented an alternative method for the simultaneous estimation of INH, PYZ and RIF in SLNs. The method fulfilled all requirements and considered as reliable, feasible and could be applied for the determination of assay, EE, in vitro drug release and stability of ATDs-loaded SLNs.

The RP-HPLC method developed was found to be a convenient and cost-effective method for the simultaneous estimation of INH, PYZ and RIF in SLNs. This method was validated according to the ICH guidelines Q2 (R1) and was found to be a reliable and feasible method, which included specificity, linearity, precision, accuracy, LOD and LOQ. The chromatographic run time was 20 min, which allowed numerous samples to be analysed in a short period, as well as reduced solvent costs and damage to the environment. Hence it was concluded that the validation parameters showed the method to be excellent for the routine quality control analysis of INH, PYZ and RIF ATDs-loaded SLNs.

TABLE 5: LINEAR REGRESSION ANALYSIS

ATDs	Linearity range (µg/ml)	Slope \pm % RSD	Intercept \pm % RSD	Correlation coefficient (R ²)
INH	8-120	34490.67 \pm 0.003967	-32035.00 \pm 0.234533	0.9980
PYZ	40-600	111140.7 \pm 0.000387	-5078.07 \pm 0.052567	0.9980
RIF	16-240	100030.3 \pm 0.001377	-101244 \pm 0.080767	0.9981

*Mean (n=3)

TABLE 6: RESULTS OF LOD AND LOQ

ATDs	SD	Slope	% RSD	LOD	LOQ
INH	43.715	34491	0.12	0.0042	0.0127
PYZ	77.797	111141	0.06	0.0023	0.0070
RIF	1439.66	100030	1.34	0.0475	0.1439

REFERENCES

- [1] Khatak S, Dureja H. Recent advances in nanotechnology based tubercular chemotherapy. *Int J Pharm Sci Nanotech* 2015;8(4):2919-34.
- [2] Global tubercular report 2015, 20th ed. World Health Organization. 2015; p. 1-115. Available from:

www.who.int/tb/publications/global_report/gtbr15_main_text.pdf.

[3] The union health solutions for the poor. International union against tuberculosis and lung disease, activity report 2012. 68, boulevard Saint-Michel-75006, Paris-France. 2012; p. 1-73. Available from: www.theunion.org/what-we-do/publications/official/english/Rapport_2012-print5LR6.pdf.

[4] Benetton SA, Kedor-Hackmann ER, Santoro MI, Borges VM. Visible spectrophotometric and first-derivative UV spectrophotometric determination of rifampicin and isoniazid in pharmaceutical preparations. *Talanta* 1998;47(3):639-43.

[5] Swamy N, Prashanth KN, Basavaiah K. Redox-reaction based spectrophotometric assay of isoniazid in pharmaceuticals. *ISRN Anal Chem* 2014;2014:717019.

[6] Kamel MS. Spectrophotometric determination of isoniazid in pure form and pharmaceutical preparation. *World J Chem* 2008;3(1):11-6.

[7] Tsai IL, Liu HY, Kuo PH, Wang JY, Shen LJ, Kuo CH. Quantitative determination of isoniazid in biological samples by cation-selective exhaustive injection-sweepingmicellar electrokinetic chromatography. *Anal Bioanal Chem* 2011;401(7):2205-14.

[8] Huang J, Zhang C, Zhang Z. Flow-injection chemiluminescence determination of isoniazid with electro-generated hypochlorite. *Fresenius J Anal Chem* 1999;363:126-28.

[9] Zheng X, Zhang Z. Flow-injection chemiluminescence determination of isoniazid using on-line electrogenerated BrO as an oxidant. *Analyst* 1999;124:763-66.

[10] Lapa RAS, Lima JLFC, Santos JLM. Fluorimetric determination of isoniazid by oxidation with cerium (IV) in a multi-commutated flow system. *Anal ChimActa* 2000;419:17-23.